DQB1*02 Product Insert Page 1 of 8

101.213-24 – including *Taq* **polymerase**General "Instructions for Use"

IFU-01 Rev. No. 00 can be downloaded from

Lot No.: **08G** Lot-specific information www.olerup.com

Olerup SSP® DQB1*02

Product number: 101.213-24 – including *Taq* polymerase

Lot number: 08G

Expiry date: 2011-August-01

Number of tests: 24 Number of Wells per test: 6

Storage - pre-aliquoted primers: dark at -20°C

- PCR Master Mix: -20°C
- Adhesive PCR seals RT
- Product Insert RT

This Product Description is only valid for Lot No. 08G.

CHANGES COMPARED TO THE PREVIOUS *OLERUP* SSP® DQB1*02 LOT

The DQB1*02 specificity and interpretation tables have been updated with the DQB1 alleles described since the previous *Olerup* SSP® DQB1*02 lot **(Lot No. 66E)** was made.

The DQB1*02 primer set is unchanged compared to the previous lot (Lot No.66E).

DQB1*02 Product Insert Page 2 of 8

101.213-24 – including *Taq* polymerase

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Lot No.: **08G** Lot-specific information www.olerup.com

PRODUCT DESCRIPTION

DQB1*02 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*0201 to DQB1*0205 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 6 PCR reactions in an 8 well PCR plate. Wells 7 to 8 are empty.

1 2 3 4 5 6 empty empty

The 8 well PCR plate is marked with 'DQ2' in silver/gray ink.

Well No. 1 is marked with the Lot No. '08G'.

The PCR plates are heat-sealed with a PCR-compatible foil.

Please note: When removing each 8 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Only DQB1 alleles will be amplified by the DQB1*02 subtyping kit. Thus, the interpretation of DQB1*02 subtypings is not influenced by the DQB2 and DQB3 genes.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*02 alleles, i.e. **DQB1*0201 to DQB1*0205**, recognized by the HLA Nomenclature Committee in July 2009¹ will give rise to unique amplification patterns by the primers in the DQB1*02 subtyping kit.

The DQB1*02 kit cannot distinguish the DQB1*020101 and DQB1*020102 alleles.

¹HLA-DQB1 alleles listed on the IMGT/HLA web page 2009-July-17, release 2.26.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

The 5 DQB1*02 alleles can be combined in 15 homozygous and heterozygous combinations. All but two of these genotypes give rise to unique amplification patterns.

CE

```
+-+-+ 0201,0204 = 0204,0204
+-+-+ 0201,0205 = 0205,0205
```

0201 = 020101 - 020102

101.213-24 – including *Taq* polymerase

General "Instructions for Use" IFU-01 Rev. No. 00 can be downloaded from

Lot No.: **08G** Lot-specific information www.olerup.com

SPECIFICITY TABLE

DQB1*02 SSP subtyping

Specificities and sizes of the PCR products of the 6 primer mixes used for DQB1*02 SSP subtyping

| Primer Mix | Size of spec. PCR product ¹ | Size of control band ² | Amplified DQB1*02 alleles | Other amplified DQB1 alleles ³ |
|-----------------------|---|-----------------------------------|--------------------------------|--|
| 1 | 120 bp | 515 bp | *020101-0202, 0204, 0205 | |
| 2 ⁴ | 85 bp | 430 bp | *0203 | |
| 3 | 145 bp | 515 bp | *020101, 020102, 0204, 0205 | *030101-0323, 0325, 040101-0403, 050101-0505, 060101-0635 |
| 4 | 140 bp | 430 bp | *0202, 0203 | |
| 5 | 145 bp | 430 bp | *0204 | |
| 6 | 180 bp | 430 bp | *0205 | |

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1*02 SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective lengths of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*02 subtyping.

In addition, well number 3 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB1 alleles only partial 3rd exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of the 3rd exon of DQB1 alleles are conserved within allelic groups.

⁴Short specific PCR fragments are less intense and not as sharp as longer specific bands.

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| INTER | INTERPRETATION TABLE | | | | | | |
|--|----------------------------------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|----------------------------------|--|
| DQB1*02 PCR-SSP subtyping | | | | | | | |
| Allele-specific amplification patterns of the 0201 to 0205 alleles | | | | | | | |
| | Well | | | | | | |
| | 1 | 2 | 3 | 4 | 5 | 6 | |
| Length of spec. | 120 | 85 | 145 | 140 | 145 | 180 | |
| PCR product | | | | | | | |
| Length of int. | 515 | 430 | 515 | 430 | 430 | 430 | |
| pos. control ¹ | | | | | | | |
| 5'-primer ² | 30(185) | 57(266) | 135(500) | 102(400) | 136(502) | 30(185) | |
| | ^{5'} -AAg ^{3'} | ^{5'} -TgA ^{3'} | ^{5'} -TgA ^{3'} | 5' -TCT 3' | ^{5'} -ACg ^{3'} | ^{5'} -AAg ^{3'} | |
| 3'-primer ³ | 57(266) | 71(309) | 170(604) | 135(500) | 170(604) | 77(326) | |
| | ^{5'} -Cgg ^{3'} | ^{5'} -CgT ^{3'} | ^{5'} -gAC ^{3'} | ^{5'} -ggC ^{3'} | ^{5'} -gAC ^{3'} | ^{5'} -CCg ^{3'} | |
| Well No. | 1 | 2 | 3 | 4 | 5 | 6 | |
| DQB1 allele ⁴ | | | | | | | |
| *020101-020102 | 1 | | 3 | | | | |
| *0202 | 1 | | | 4 | | | |
| *0203 | | 2 | | 4 | | | |
| *0204 | 1 | | 3 | | 5 | | |
| *0205 | 1 | | 3 | | | 6 | |
| *030101-0323, 0325, | | | | | | | |
| 040101-0403, 050101- | | | 3 | | | | |
| 0505, 060101-0635 | | | | | | | |
| DQB1 allele ⁴ | | | | | | _ | |
| Well No. | 1 | 2 | 3 | 4 | 5 | 6 | |

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*02 subtyping.

In addition, well number 3 contains the primer pair giving rise to the longer, 515 bp, internal

positive control band in order to allow kit identification. ²The codon, and in parenthesis the nucleotide, in the 2nd or 3rd exon, matching the specificitydetermining 3'-end of the primer is given. Codon and nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is

given. Empty spaces indicate codon boundaries. ³The codon, and in parenthesis the nucleotide, in the 2nd or 3rd exon, matching the specificitydetermining 3'-end of the primer is given in the anti-sense direction. Codon and nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal

nucleotides of the primer is given. Empty spaces indicate codon boundaries.

⁴For several DQB1 alleles only partial 3rd exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of the 3rd exon of DQB1 alleles are conserved within allelic groups.

Lot No.: **08G** Lot-specific information www.olerup.com

| CELL LINE VALIDATION SHEET | | | | | | | | | | |
|----------------------------|------|-------------|-------|-------------------|------------|-----------|-----------|-----------|-----------|-----------|
| DQB1*02 SSP subtyping kit | | | | | | | | | | |
| | | | | | | | W | ell | | |
| | | | | | 1 | 2 | 3 | 4 | 5 | 6 |
| | | | | | 7 | Ø | 3 | 4 | Ω | စ္ |
| | | | | e G | 200961701 | 200961702 | 200961703 | 200961704 | 200961705 | 200961706 |
| | | | | Production No. | 61 | 19 | 19 | 19 | 19 | 19 |
| | | | | od . | 90 | 000 | 000 | 000 | 000 | 000 |
| | | | | P S | 20 | 20 | 20 | 20 | 20 | 20 |
| | IHW | C cell line | D | QB1 | | | | | | |
| 1 | 9001 | SA | *0501 | | - | - | + | - | - | - |
| 2 | 9280 | LK707 | *0601 | *0202 | + | - | + | + | - | - |
| 3 | 9011 | E4181324 | *0601 | | - | - | + | - | - | - |
| 4 | 9275 | GU373 | *0201 | | + | - | + | - | - | - |
| 5 | 9009 | KAS011 | *0502 | | - | - | + | - | - | - |
| 6 | 9353 | SM | *0302 | *0601 | - | - | + | - | - | - |
| 7 | 9020 | QBL | *0201 | | + | - | + | - | - | - |
| 8 | 9025 | DEU | *0301 | | - | - | + | - | - | - |
| 9 | 9026 | | *0302 | | - | - | + | - | - | - |
| 10 | | LKT3 | *0401 | | - | - | + | - | - | - |
| 11 | 9051 | PITOUT | *0202 | | + | - | - | + | - | - |
| 12 | 9052 | DBB | *0303 | | - | - | + | - | - | - |
| 13 | | JESTHOM | *0501 | | - | - | + | - | - | - |
| 14 | 9071 | OLGA | *0402 | | - | - | + | - | - | - |
| 15 | 9075 | | *0303 | | - | - | + | - | - | - |
| 16 | 9037 | SWEIG007 | *0301 | | - | - | + | - | - | - |
| 17 | | CTM3953540 | *0201 | *0603 | + | - | + | - | - | - |
| 18 | 9257 | 32367 | *0602 | *0202 | + | - | + | + | - | - |
| 19 | | BM16 | *0301 | | - | - | + | - | - | - |
| 20 | 9059 | | *0604 | | - | - | + | - | - | - |
| 21 | | AMALA | *0301 | | - | - | + | - | - | - |
| 22 | | KOSE | *0503 | *0604 | - | - | + | - | - | - |
| 23 | 9124 | | *0503 | *0601 | - | - | + | - | - | - |
| 24 | 9035 | | *0301 | | - | - | + | - | - | - |
| 25 | | IBW9 | *0202 | | + | - | - | + | - | _ |
| 26 | | WT49 | *0201 | | ÷ | - | + | Ė | - | _ |
| 27 | 9191 | CH1007 | *0401 | *0501 | Ė | - | + | - | - | _ |
| 28 | | BEL5GB | *0202 | *0301 | + | - | + | + | - | _ |
| 29 | 9050 | | *0202 | 0001 | ÷ | - | - | + | - | _ |
| 30 | 9021 | | *0402 | | Ė | - | + | - | - | _ |
| 31 | | DUCAF | *0201 | | + | - | + | - | - | - |
| 32 | | HAG | *0301 | | - | - | + | - | - | - |
| 33 | | MT14B | *0302 | | - | - | + | - | - | _ |
| 34 | 9104 | | *0301 | | - | - | + | - | - | - |
| 35 | | SSTO | *0305 | | - | - | + | - | - | - |
| 36 | | KT17 | *0302 | 1 | - | - | + | - | - | - |
| 37 | | HHKB | *0603 | 1 | - | - | + | - | - | - |
| 38 | 9099 | | *0301 | | - | - | + | - | - | - |
| 39 | 9315 | | *0201 | *0301 | + | - | + | - | - | - |
| 40 | | WHONP199 | *0202 | *0303 | ÷ | - | + | + | - | - |
| 41 | | H0301 | *0609 | 1000 | Ė | - | + | ÷ | - | - |
| 42 | | TAB089 | *0601 | | - | - | + | - | - | - |
| 43 | | T7526 | *0303 | | - | - | + | - | - | - |
| 44 | 9057 | | *0503 | | - | - | + | - | - | _ |
| 45 | | SHJO | *0202 | | + | - | Ė | + | - | _ |
| 46 | | SCHU | *0602 | | Ė | - | + | - | - | _ |
| 47 | | TUBO | *0301 | | <u> </u> | i. | + | - | - | - |
| 48 | | TER-ND | *0501 | | | - | + | | _ | _ |
| 40 | 9303 | I LIK-IND | 0301 | | | | | - | | |

DQB1*02 Product Insert Page 6 of 8

101.213-24 – including *Taq* polymerase

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Lot No.: **08G** Lot-specific information www.olerup.com

CERTIFICATE OF ANALYSIS

Olerup SSP® DQB1*02 SSP

Product number: 101.213-24 – including *Taq* polymerase

Lot number: 08G

Expiry date: 2011-August-01

Number of tests: 24 Number of wells per test: 6

Well specifications:

| Well No. | Production No. |
|----------|----------------|
| 1 | 2009-617-01 |
| 2 | 2009-617-02 |
| 3 | 2009-617-03 |
| 4 | 2009-617-04 |
| 5 | 2009-617-05 |
| 6 | 2009-617-06 |

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 2, 5 and 6 were available. The specificities of the primers in primer solutions 2 and 6 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer. In primer solution 5 it was only possible to test the 3'-primer, the 5'-primer was not possible to test.

Results: No false positive or false negative amplifications were obtained.

Date of approval: 2009-August-25

Approved by:

Quality Control, Supervisor

DQB1*02 Product Insert Page 7 of 8 101.213-24 – including *Taq* polymerase General "Instructions for Use"

IFU-01 Rev. No. 00 can be downloaded from

Lot No.: **08G** Lot-specific information www.olerup.com

Declaration of Conformity

Product name: Olerup SSP® DQB1*02

Product number: 101.213-24

Lot number: 08G

Intended use: DQB1*02 high resolution histocompatibility testing

Manufacturer: Olerup SSP AB

Hasselstigen 1

SE-133 33 Saltsjöbaden, Sweden

Phone: +46-8-717 88 27 **Fax:** +46-8-717 88 18

We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2000 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex III, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden 2009-August-25

Olle Olerup Managing Director

August 2009

Rev. No.: 00

DQB1*02 Product Insert Page 8 of 8 101.213-24 – including *Taq* polymerase General "Instructions for Use"

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